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**Research Article** 

# Nephroprotective and Anti-Inflammatory Activity of Combination of Punica Granatum (Fruit Peel) And Tectona Grandis (Bark)

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### ABSTRACT

Herbal medication in addition referred to as herbal treatment or phytoconstuients refers to employ a plant's seed, berries, roots, leaves, bark or flowers for healthful functions. The procedure of herbs to treat a variety of different ailments is universal and exists in every human culture on Earth. The plants Punica granatum and Tectona grandis is important medicinally and claimed to be helpful in the treatment of different ailments like diabetes, inflammations, eruptions and bites of poisonous animals etc., and contains different classes of compounds of pharmacological importance such as, alkaloids, glycosides, saponins, flavonoids and triterpines and have been screen biologically for Nephroprotective and anti-inflammatory actions.

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### INTRODUCTION

Herbal medication in addition referred to as herbal treatment or phytomedicine refers to, berries, roots, leaves, bark or flowers for healthful functions<sup>1-5</sup>. Herbalist contains a long tradition of use outside of typical medication. Ancient Chinese and Egyptian papyrus writings describe healthy use for plants at the same as early as  $3000 \text{ BC}^3$ . Native cultures (such as African and Native American) use herbs in their heal rituals, whereas others developed ancient healthy systems (such as written material and ancient Chinese medicine) during which seasoned therapy were used. Researchers start that folks in numerous elements of the globe attend uses of similar plants for a similar purpose<sup>6-9</sup>. The procedure of herbs to treat a variety of different ailments is universal and exists in every human culture on Earth. Despite this, the major use of medical herbs still occurs in societies which are not fully developed Because of the high costs involved with industrialized modern medicines, many people living in increasing nations simply do not have the financial resources to give for them, and as a result, they are enforced to use normal herbs as an affordable alternative.<sup>10-</sup> 14



Fig.No.1 Herbs

The plants Punica granatum and Tectona grandis is important medicinally and claimed to be helpful in the treatment of different ailments like diabetes, inflammations, eruptions and bites of poisonous animals etc., and contains different classes of compounds of pharmacological importance such as, alkaloids, glycosides, saponins, flavonoids and triterpines and have been screen biologically for Nephroprotective and anti-inflammatory actions<sup>15-</sup> <sup>18</sup>.



Fig. No.: 2 Fruit peel of Punica granatum



Fig. No.: 3 Tectona grandis plant

### **METERIALS AND METHOD**

Table No: 1 List of materials and equipment's usedduring experiment.

Sr. No	Name of the materials and equipment			
1	Anesthetic ether (Sigma Solvents and			
1	Pharmaceuticals, Mumbai)			
2	Formalin (Hi-Media Laboratory, Mumbai.)			
3	Ethanol (Nice-Cochin)			
4	Carrageenin (Nice-Cochin)			
5	Diclofenac sodium (Cipla, Mumbai)			
6	Distilled water.			
7	Evans blue (S.D. Fine chemicals, Mumbai)			
0	Lead Mercury (S.D. Fine chemicals,			
0	Mumbai)			
0	Restraint cages (Shri. Venkateswara			
<sup>9</sup> Enterprises, Bangalore)				
10	Chloroform (Nice-Cochin)			
11	Refrigerator (Whirlpool-Kelvinator India			
11	Ltd, India)			



# Collection and authentification of plant materials:

The stem of *Punica granatum and Tectona grandis* were collected from limited area and authentified by botanist Dr. Narasingham BSI.

# Preparation of special extracts (petrolium ether, chloroform, ethanolic and aqueous extracts):

The fruit peel of *Punica granatum* and bark of Tectona grandis were dried in shade at room high temperature then Subjected to size decrease to a fine powder with the help of mixer grinder. Then the powder was extract with petroleum ether, chloroform, ethanol and water consecutively by simple extraction. The extract was move into the before weighed clear beaker and evaporate to a thick paste on the water bath, maintained at 50°C to get extracts. The extract be lastly air dry thoroughly to remove all traces of the solvent and the percentage yield was planned each time before extracting with next solvent, marc will be dried in hot air oven below 50°C and each extract will be concentrated by distilling off the solvent and then evaporated to dryness on a water bath to get the extracts.

## Pharmacological evaluation:

Depending upon the occurrence of phytoconstituent in the different extracts, the ethanolic and aqueous extracts are selected for the following pharmacological actions.

- Anti-inflammatory activity
- Nephroprotective activity

# Anti-Inflammatory Activity By Carrageenan Induced Paw Oedema<sup>60, 61,74</sup>

Group A: Control (Carrageenin 1%) Group B: Standard (Diclofenac 30 mg/ kg) Group C: EEPG+EETG (200 mg/kg p.o) Group D: EEPG+EETG (400 mg/kg p.o) Group E: AEPG+AETG (200 mg/kg p.o) Group F: AEPG+AETG (400 mg/kg p.o)

## **Experimental Procedure:**

Albino rats (150-200 g) be separated into 6 groups each containing 6 animals they be fast overnight prior to and through the test but include free access to water. Group A be served as toxicant control treated among toxicant Carrageenan, group B with Diclofenac (40 mg/kg p.o.) that serve because normal Groups C, D, E and F be administer with EEPG, EETG and AEPG, AETG (200 mg/kg and 400 mg/kg dose p.o) respectively. The rats of groups B, C, D, E and F be administer with 1% of Carrageenin into sub plantar part of right hind paw after administration of rats 1h of Diclofenac/extracts. directly here later than the oedema volumes of the injected paws be considered plethysmographically at prefixed time interval. The variation among paw volume of the treat animals is considered and the mean oedema quantity be calculated19-28. Percentage decrease in oedema volume was calculated by use the formula.

Percentage reduction = 
$$\frac{\text{Vo - Vt}}{\text{Vo}} \ge 100$$

Where,

Vo = Volume of the paw of control at time 't'. Vt = Volume of the paw of drug treated at time 't'. Statistical analysis:

All consequences will be expressed as mean  $\pm$  SEM from 6 animals. Statistical variation in mean will be analyze use one-way ANOVA (analysis of variance) follow by position hoc test (Dunnett's't' test). P< 0.05\*, 0.01\*\* and 0.001\*\*\* will be measured as statistically important<sup>29-37</sup>.

# Nephroprotective activity by Gentamicin Induced Nephrotoxicity in Rats<sup>47, 52</sup>

The evaluation of the ethanolic extract for nephroprotective action be done according to the process known in the literature with minor modification.



Total 36 animals are in use and 6 rats were selected in each of the follow groups:

Group I: Control group

Group II: Gentamicin control group (60mg/kg)

Group III: Gentamicin + Ethanolic extract

(400mg/kg)

Group IV: Gentamicin + ethanolic extract (200mg/kg)

Group V: Gentamicin + Aqueous extract (400mg/kg)

Group IV: Gentamicin + Aqueous extract (200mg/kg).

# Activity profile of the test formulations in gentamicin induced changes in different parameters:

Serum uric acid is the ending product of purine catabolism. So, any fault in the glomerular filtration rate causes the rise into the stage of uric acid in the blood. The raise later than gentamicin can be credited to the GFR impairment. The exchange of the elevation by any substance may be analytic of the exchange of the GFR impairment53-59.

## **RESULTS AND DISCUSSION**

The nature and percentage yield of different extract be given below:

# Table No.: 2 Nature and Percentage yield of the extracts

Sr. No.	Name of the Extract	Nature	Colour	%Yield (w/w) g
1	Pet ether	Sticky	Dark	2.00
1.	I et.ether	Sticky	green	2.00
2	Chloroform	Sticky	Dark	6 50
۷.	Chiofololini	Sticky	green	0.50
3	Ethanol	Sticky	Dark	9.50
5	Ethanoi	Sticky	green	9.50
4		Sticky	Dark	11.00
+	Aqueous	SUCKY	brown	11.00

### Preliminary phytochemical screening:

EEPG+EETG and AEPG+AETG were subjected for phytochemical selection and start to include tannins, sterols, flavonoids, glycosides, and alkaloids triterpines in ethanolic and aqueous extracts. The phytochemical constituents of different extract of Punica granatum and Tectona grandis are shown below:

Table No.: 3. Phytochemical evaluation of combined extracts of *Punica granatum* and *Tectona* 

grandis

8					
Sr. No.	Tests	Petroleu mether	Chloro form	Etha nol	Water
01	Alkaloids	-ve	-ve	+ve	+ve
02	Carbohydrates	-ve	-ve	+ve	+ve
03	Flavonoids	-ve	-ve	+ve	+ve
04	Saponins	-ve	-ve	+ve	+ve
05	Sterols	+ve	+ve	+ve	+ve
06	Tannins	-ve	-ve	+ve	+ve
07	Glycosides	-ve	-ve	+ve	+ve

# Pharmacological activities: Acute oral toxicity study:

The mice treated with EEPG+EETG and AEPG+AETG at a dose of 200 mg/kg, p.o. exhibited usual behaviour, without any symbols of passivity, stereotypy and vocalization. Their motor movement and secretory symbols were also usual and no sign of depression. EEPG+EETG and AEPG+AETG even up to the dose level of 2000 mg/kg body weight did not create any behavioural symptoms or mortality. So 1/10th and 1/5th doses of (maximum dose tested for each extract) be chosen as medium and high dose and were used in the current study to investigate nephroprotective and anti- inflammatory actions

# Anti-inflammatory activity by Carrageenin induced paw oedema model in rats:

The EEPG+EETG and AEPG+AETG with the selected doses i.e. 200 and 400 mg/kg consist of show a important drop in paw oedema volume in



Carrageenin induced paw oedema in rats at different time interval. Results are tabulated in Table No. 4.7. Diclofenac sodium (30 mg/kg) was used as typical situation and it has considerably reduced paw oedema volume by 17.81% at 1st h, 27.58% at 2nd h, 50.09% at 3rd h and74.13 % at 4th h, which was created to be a moment dependent result.

During 1st h of study EEPG+EETG and AEPG+AETG with 200mg/kg and 400 mg/kg doses have extensively reduced oedema volume by 3.43, 10.72, 3.43, 7.72 % respectively noted as time dependent result.

During 2nd h of study EEPG+EETG and record as time dependent effect and result be AEPG+AETG with 200mg/kg and 400 mg/kg graphically represent in Fig No.:4.5. **Table No. 4. Anti-inflammatory effect of EEPG+EETG and AEPG+AETG on paw volume in Carrageenin** 

dose have significantly reduced oedema volume by 13.9, 24.28, 15.11, 17.29.

% respectively noted as time reliant result.

During 3rd h of study EEPG+EETG and AEPG+AETG with 200mg/kg and 400 mg/kg doses have considerably reduced oedema volume by 32.07, 34.33, 38.08,

47.46 % correspondingly noted as time dependent result.

During 4th h of study EEPG+EETG and AEPG+AETG with medium and high doses have significantly reduced oedema volume 51.72, 56.89, 55.17, 60.34 % respectively which was record as time dependent effect and result be graphically represent in Fig No.:4.5.

induced paw edema in rats						
Crowns	Tuestment	Paw oedema volume				
Groups	I reatment	60 min	120 min	180 min	240 min	
Control	Carragenin	$0.466 \pm 0.042$	$0.4833 \pm 0.047$	$0.533 \pm 0.04$	$0.58 \pm 0.030$	
Standard	Diclofenac	$0.383 \pm 0.030$	0.35±0.02236	$0.266 \pm 0.033$	0.15±0.0223	
	sodium				6	
	(30mg/kg)					
EEPG+EETG	200mg/kg	$0.45 \pm 0.0428$	$0.4166 \pm 0.047$	$0.366 \pm 0.033$	$0.2833 \pm 0.03$	
					0	
EEPG+EETG	400mg/kg	$0.416\pm0.047$	0.366±0.033	$0.35 \pm 0.022$	$0.25 \pm 0.022$	
AEPG+AETG	200mg/kg	0.45±0.0428	0.41±0.036	0.33±0.0421	0.26±0.049	
AEPG+AETG	400mg/kg	0.43±0.0210	0.4±0.042	$0.28 \pm 0.030$	0.233±0.033	



Groups

Fig. No. 4 Anti-inflammatory effect of EEPG+EETG and AEPG+AETG on pawvolume in Carrageenin induced paw oedema in rats



Croups	Treatmont	% Reduction in paw volume			
Groups	Treatment	60 min	120 min	180 min	240 min
Control	Carragenin	-	-	-	-
Standard	Diclofenac sodium	17.81	27.58	50.09	74.13
	(30mg/kg)				
EEPG+EETG	200mg/kg	3.43	13.9	32.07	51.72
EEPG+EETG	400mg/kg	10.72	24.28	34.33	56.89
AEPG+AETG	200mg/kg	3.43	15.11	38.08	55.17
AEPG+AETG	400mg/kg	7.72	17.29	47.46	60.34

Table No. 5. Percentage reduction of paw volume in Carrageenin induced paw oedema



Fig. No. 5 Percentage reduction of Paw oedema volume in different groups

Nephroprotective activity by Gentamicin induced nephrotoxicity in albino rat's model:

Gentamicin like other aminoglycoside antibiotics causes nephrotoxicity by inhibiting protein synthesis in renal cells. This alters the body weight and elevates the usual levels of blood urea and serum creatinine.

The administration of ethanolic and aqueous extracts of combined doses (200mg/kg and 400 mg/kg) of Punica granatum and Tectona grandis has shown a significant reduction in blood urea and serum creatinine levels as shown in the tables below.

Table No: 6 % of Body weight change in	
gentamicin induced Nephrotoxicity in albino ra	ts

Crouns	% of body weight	
Groups	change	
Normal control	$3.44\pm0.190$	
Toxic control	$9.926 \pm 0.448 ***$	
EEPG+EETG (200mg/kg)	$8.216 \pm 0.410^{\textit{***}}$	
EEPG+EETG (400mg/kg)	$5.776 \pm 0.463 **$	
AEPG+AETG (200mg/kg)	$6.3 \pm 0.44$	
AEPG+AETG (400mg/kg)	$4.4 \pm 0.411$	

Values are expressed in mean  $\pm$ SEM where n = 6, Significant at P < 0.05\*, 0.01\*\* and 0.001\*\*\*, compared to control group.





Groups

Fig. No: 6 % of Body weight changes in special groups by Gentamycin inducedNephrotoxicity

#### model.

Table No.: 7 Blood urea and serum creatinine levels of different groups by gentamicin induced Nephrotoxicity

(cpiii otomicity)				
Groups	Blood urea	Serum creatinine		
Normal control	$31.03 \pm 5.018$	$1.023{\pm}~0.053$		
Toxic control	$70.591 \pm 4.064 \text{***}$	$2.05{\pm}~0.081{}^{\boldsymbol{\ast\ast\ast\ast}}$		
EEPG+EETG (200mg/kg)	57.385 ± 11.724**	1.918± 0.07***		
EEPG+EETG (400mg/kg)	45.916 ± 11.65*	1.53 ± 0.11***		
AEPG+AETG (200mg/kg)	51.948 ± 3.673***	1.668± 0.080***		
AEPG+AETG (400mg/kg)	$38.046\pm5.280$	$1.191 \pm 0.053$		

Values are expressed in mean  $\pm$ SEM where n = 6, Significant at P < 0.05\*, 0.01\*\* and 0.001\*\*\*, compared to control group.



Groups

Fig. No:7. Blood urea level of different groups in gentamicin induced Nephrotoxicity model

Serum creatinine (mg/dl) of different groups



Fig. No: 8. Serum creatinine level of special groups in gentamicin induced Nephrotoxicitymodel DISCUSSION

In the current study the fruit peel of Punica granatum and bark of Tectona grandis were chosen the anti-inflammatory activity. It was for previously reported that the individuals of the Punica granatum and Tectona grandis have potent anti-inflammatory activity. The preliminary phytochemical research reveals the presence of alkaloids, flavonoids, saponins, tannins and glycosides in both ethanolic and aqueous extracts. Inflammation is a state which is cause by chemical mediators like prostaglandins and cytokinin's. It is report that the phytochemicals like flavonoids, saponins and tannins decrease the inflammation. In the current study the above-mentioned phytochemicals are current in both the extracts this can be recognized for its anti- inflammatory movement.

The fruit peel Punica granatum be reported to contain phenolic compounds like flavonoids, which possess antioxidant property, The fruit peel of Punica granatum were previously report for its nephroprotective movement on diabetes-induced renal damage. Nephrotoxicity is a regular cause of human being which is mainly caused due to oxidative stress. In the current study both the extracts containing polyphenolic compounds have antioxidant property, so the nephroprotective activity of the above-mentioned plants Punica



granatum and Tectona grandis is appropriate to its antioxidant property.

## CONCLUSION

In the present study Punica granatum and Tectona grandis both the extracts significantly reduced the nephrotoxicity and inflammation. Punica granatum and Tectona grandis both nephrotoxicity and inflammation may be due to creation of inflammatory mediators by oxidative stress. In the present study is observed that due to the presence of phytoconstituents like alkaloids, flavonoids, saponins and tannins which also decrease the oxidative stress and inflammatory mediators. The anti-inflammatory and nephroprotective activity may be dependable for above phytoconstituents.

Further studies are essential to begin the exact mechanism of phytoconstituents responsible for the above actions.

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## **CONFLICT OF INTEREST**

The authors state that there is NO conflict of interest.

# REFERENCES

- Christopher EC, Elizabeth A, Toby JL, Taixiang WU; Herbal interventions for chronic asthma in adults and children: A methodical Review and Meta-Analysis; Primary Care Respiratory Journal ;2010;19(4);307-314.
- Seyyed SA; greatest Treatment for Allergic Asthma among Traditional Herbal Medicine: A Brief Report; American-Eurasian Journal of Agriculture and Environmental Sciences;2013;13(2);291-292.
- Munge B, Sreedhar G, Mounika Y, Suddagoni
   S. Anthelmintic activity of seaweed extract of Padina boergesenii and Padina tetrastromatica

acute subacute level in naturally infected sheep. Natl J Physiol Pharm Pharmacol 2022;12(02):115-120

- Edward M, Curtis C, Dugald S and Izzy K; African herbal medicines in the treatment of HIV: Hypoxis and Sutherlandia. An summary of data and pharmacology; Nutrition Journal 2005; 4(19);1-6.
- Kavishankar GB, Lakshmide N, Mahadeva MS, Prakash HS and Niranjana SR; Diabetes and medicinal plants-A review; International Journal of Pharmacy and Biomed Sciences; 2011, 2(3), 65-80.
- Gautam AH, Sharma R and Rana AC; Review on Herbal Plants Usefully in Tuberculosis; International Research Journal Of Pharmacy; 2012;3(7);64-67.
- 7) Prachi G, Haruyo I, Nikita M, Gautam S and Bharat BA; From Ancient Medicine to recent Medicine: Ayurvedic concept of Health and Their Role in Inflammation and Cancer; Journal of the Society for Integrative Oncology; 2007;5(1);1-16.
- 8) WHO procedure for assess excellence of herbal medicine with reference to contaminants And residues 2007.
- Sheetal V and Singh SP; Current and impending position of herbal medicines; Veterinary World; 2008;1(11);347-350.
- 10) Sanjoy KP and Yogeshwer S; Herbal Medicine: present place and the Future; Asian Pacific Journal of Cancer Prevention; 2003( 4);281-288.
- 11) Sandhya W; Global healthiness care dare: Indian experience and new prescriptions; Electronic Journal of Biotechnology; 2004;7(3);217-223.
- 12) Harish KH, Prashanth KJ and Shruthi SD; Pharmacognostic and Phytochemical studies



on the undergrowth of Murraya koenigii (l) spreng ; Pharmacophore; 2010;1(3);231-238.

- 13) Kawaljeet, Arvind KA, Sayeed A and Perwez A; Pharmacognostic study on bark of Murraya koenigii Spreng; International Journal of Research in Pharmaceutical and Biomedical Sciences; 2011;2(4);1670-1677.
- 14) Haslizawati abu BN,Aspollah S, Mawardi R, Sharif1 A, Khalid K and Umi KY; element constituent from stem barks and extraction of Murraya koenigii (rutaceae); The Malaysian Journal Of Analytical Sciences; 2007;11(1); 173-176.
- 15) Abishek I, Sunil P, Hemanth P and Lindsay B; Potential health repayment of Indian spices in the symptom of the medical syndrome : A review; Indian Journal of Biochemistry and Biophysics; 46(6);467-481.
- 16) Ajay S, Rahul S, Sumit G, Paras M, Mishra A and Gaurav; inclusive review:Murraya koenigii Linn; Asian Journal of Pharmacy and Life Science; 2011;1(4);417-425.
- 17) Kokate CK, Purohit AP, Gokhale SB. common Introduction. Text book of Pharmacognosy. 20th ed. Pune: Nirali Prakashan; 1996:1.
- 18) Rang HP, Dale MM, Ritter JM, Moore PK. Drugs affecting major organ systems. Text volume of Pharmacology. 5th ed. UK: Elsevier science Limited; 2003: 368.
- 19) Carin ED and priya G; Nonsteroidal antiinflammatory drugs; Physical Medicine And Rehabilation Clinics Of North America; 2006(17);347-354.
- 20) Rainsford KD; Anti-inflammatory drugs inside the 21st century; Inflammation in the pathogenesis of chronic diseases; 2007; 3-27.
- 21) Aruna D and Thirunethiran K; Evaluation of Anti-inflammatory activity and analgesic action of Aloe vera leaf take out in rats;

International Research Journal Of Pharmacy; 2011; 2(3); 103-110.

- 22) Langmead L, Makins RJ, Rampton DS; Antiinflammatory property of Aloe vera gel in Human Colorectal Mucosa in vitro ; Aliment Pharmacol Ther 2004;(19); 521–527.
- 23) Vaibhavi J, Rakesh, Pankaj K, Neeraj P, Sunil G, Anupriya P and Sonu S; Cinnamon: A Pharmacological evaluation; Journal of Advanced Scientific Research; 2010;1(2);19-23.
- 24) Rajesh KM, Anil K and Ashok K; pharmacological movement of Zingiber officinale; International Journal Of Pharmaceutical and Chemical Sciences; 2012;1(3);1073-1078.
- 25) Vijay SJ,Santosh KS, Pankaj K and Ashish KS ; modern Pharmacological Trends of Glycyrrhiza glabra Linn; International Journal of Pharmaceutical Frontier Research; 2011; 1(1):170-185.
- 26) Sharma P, Tomar L, Bachwani M and Bansal V; Review on Neem (Azadirachta indica):Thousand problems one solution; International Research Journal Of Pharmacy; 2011;2(12);97-102.
- 27) Girish K and Shankara BS; Neem A Green Treasure; Electronic Journal of Biology;2008;4(3);102-111.
- 28) Baburao B, Rajyalakshmi G, Venkatesham A, Kiranb G, Shyam SA and Ganga BR antiinflammatory and antimicrobial actions of methanolic extract of Tribulus terrestris linn place; International Journal Of Chemical Science; 7(3), 2009, 1867-1872.
- 29) Singh G, Sharma PK, Dudhe R and Singh S; Biological actions of Withania somnifera; Annals of Biological Research; 2010;1; (3):56-63.



- 30) Veena S, Sadhana S, Pracheta and Ritu P; Withania somnifera: A rejuvenate Ayurvedic Medicinal Herb for the Treatment of a variety of Human ailments; International Journal of PharmTech Research; 2011;3(1);187-192.
- 31) Lakshmi CM, Betsy BS, Simon D; Scientific foundation for the Therapeutic Use of Withania somnifera (Ashwagandha):A Review; Alternative Medicine Review; 2000;5(4);334-346.
- 32) Biplab B and Jhinuk C; Identification of Proapoptopic, Anti-Inflammatory, Anti-Proliferative, Anti-Invasive and Anti-Angiogenic Targets of Essential Oils in Cardamom by Dual Reverse Virtual Screening and Binding Pose Analysis; Asian Pacific Journal of Cancer Prevention; 2013;14(6);3735-3742.
- 33) Shashank M, Ajay KJ, Manoj J, Cathrin M and Debjit B; Analgesic And Anti-Inflammatory action of Kalanchoe pinnata (Lam.) Pers; Journal of Medicinal Plants Studies 2013;1(2);24-28.
- 34) Jothibai RM, Kumaresan S and Ravikumar S; A preliminary study on the anti- inflammatory action of Methanol take out of Ulva lactuca in rat; Journal of Environmental Biology; 2009;30(5);899-902.
- 35) Sabiha S, Aftab AM, Asif M and Akhtar M; Myrtus communis Linn-A review Indian Journal of Natural goods and Resources; 2011;2(4),395-402.
- 36) Sekhar S, Karmakar R, Ramachandra KK, Ramachandrappa SN and Harischandra SP; Potential Anti-inflammatory Bioactives from Medicinal undergrowth of western ghats, India; Pharmacognosy communications; 2012;2(2);1-5.37)Fawzi I, Kamal M, Ahmad BK and Talal A; Hepatoprotetive, Cardioprotective and Nephroprotective

Actions of Essential Oil Extract of Artemisia sieberi in

- 37) Alloxan Induced Diabetic Rats; Iranian Journal of Pharmaceutical Research (2012);11(4);1227-1234.
- 38) Prusty KB, Harish B and Mamatha CH; Evaluation of Nephroprotective Activity of the Methanolic Extract of Leaves of Bauhinia variegata Linn, (Family- Caesalpiniaceae); Journal of PharmaSciTech; 2012; 2(1);16-19.
- 39) Cordeiro MC and kaliwal BB;Hepatoprotective and Nephroprotective activity of bark take out of Bridelia retusa spreng in Ccl4 treat female mice; International Journal of Molecular Biology; 2011;2(1);22-30.
- 40) Mahgoub M and Ahmed; Biochemical Studies on Nephroprotective Effect of Carob (Ceratonia siliqua L.) Growing in Egypt; Nature and Science; 2010;8(3);41-47.
- Murthy RLN, Nataraj HN and Ramachandra SS; Nephroprotective Activity of Cyanotis fasciculata next to Cisplatin in duced nephrotoxicity; International Research Journal Of Pharmacy; 2011;2(9);137-142.
- 42) Sahoo HB, Swain SR, Nandy S, Sagar R and Bhaiji A; Nephroprotective action Of Elephantophus scaber leaves; International Research Journal of Pharmacy;2012;3(5);246-250.
- 43) Shivalinge KP and Vrushabendra BM; Histopathological and Nephroprotective study of aqueous stem bark take out of Ficus racemosa in drug induced nephrotoxic rats; International Organization of Scientific Research Journal of Pharmacy; 2012; 2(2) 265-270.
- 44) Shivalinge KP and Vrushabendra BM; Study of nephroprotective activities of stem bark extract of Ficus racemosa in gentamicin



induced sharp renal failure in rats International Journal Of Pharmaceutical and Chemical Sciences; 20121(1);314-318.

- 45) Saifuddin K, Hakeemuddin K, Aaftab A and Shahid A; Nephroprotective effect of the ethanolic take out of Lantana camara Linn flower taking place acute dose of Cisplatin induced renal injured rats; Rajiv Gandhi university of Health Sciences Journal of Pharmaceutical Sciences; 2012;2(2);68-77.
- 46) Bharti D, Talele, Raghunath T, Mahajan, Manojkumar Z, Chopda and Namrata VN; Nephroprotective plants: a review; International Journal of Pharmacy and Pharmaceutical Sciences; 2012;4(1);8-16.
- 47) Kanchan G, Pradeep D, Pushpalata C, Joshi YM and Vilasrao K; A re-evaluate on some Nephroprotective Medicinal vegetation; International Journal of Pharmaceutical Sciences and Research; 2012;3(8); 2451-2454.
- 48) Sarvankumar G, Lalitha V, Singaravel S, Sharif SH and Thangavel S; Nephroprotective action of Vitex negundo linn bark next to chemical induce toxicity in experimenal rats; pharmanest - An International Journal of Advances in Pharmaceutical Sciences; 2011;2(5-6);462-470.
- 49) Mehul VM, Nilesh MP, Dharmesh ND, Sarav AD, and Bhaskar VH; Assessment of Nephroprotective Potential of Sida cordifolia linn. in new animals; Scholars Research Library; 2012, 4 (1):175-180.
- 50) Olagunjua JA, Adeneyeb AA, Fagbohunkac BS, Bisugac NA, Ketikuc AO, Benebod AS, Olufowobic OM, Adeoyec AG, Alimic MA and Adeleke AG; Nephroprotective performance of the aqueous seed extract of Carica papaya linn. in carbon tetrachloride induce renal offended wistar rats: a dose- and

time-dependent relative study; Biology and medicine; 2009;1(1);11-19.

- 51) Fahima FK, Saleh IA, Shaimaa MS, Adnan AB, Nagwa SE, Abdallah I, Nabil AS and Maged SA; assessment of the Hepatoprotective, Nephroprotective and Anti-Malarial actions of different parts of Bauhinia purported and Tipuana speciosa grown in Egypt; Journal of Medicinal Plants Research; 2013;7(17);1190-1200.
- 52) Kannappan N, Madhukar A, Mariymmal, Uma sindhura P and Mannavalan R; Evaluation of nephroprotective action of Orthosiphon stamineus benth extract using rat model International Journal of Pharmtech Research;2010;2(1),209-215.
- 53) Uma C, Poornima K, Surya S, Ravikumar G and Gopalakrishnan VK; Nephroprotective Effect of Ethanolic Extract of Tabernaemontana coronaria in Mercuric Chloride induce Renal injure in Wistar Albino Rats; International Journal of Chemical Engineering and Application; 2012;3(4);269-273.
- 54) Khuntia TK, panda DS; Evaluation of Antibacterial, Antifungal and Anthelmintic action of Murraya koenigii spreng. pharma science observe an International Journal of Pharmaceutical Sciences ;2011;2(2); 0976-7908.
- 55) Dinesh KP; B; Anti-ulcer action of aqueous extract of Murraya koenigii in albino rats; International Journal of Pharma and Bio Sciences; 2011; 2 (1)524-529.
- 56) Mishra MK, Sahu RV, Mahesh G, Narendra P and Kailash P; Anti-fungal potential of sheet extract of Murraya koenigii; International Journal Of Research in Ayurveda & Pharmacy; 2010;1(2);549-552.



- 57) Bhaskaran C, Ratha B, kanimozi D;Screening for antimicriobial activity and phytochemical investigation of a variety of leaf extract of Murraya koenigii ; International Journal of Research in Ayurveda and Pharmacy;2011; 2(6); 1807-1810
- 58) Abhishek M; Anti-inflammatory action of plants extracts of Murraya koenigii. International Journal of Pharma and Bio Sciences; 2011;2(1);541-544.
- 59) Praveen S, Vidyasagar G, Anil B, Sunder S, Santosh G, Ashish A, Swapnil G and Mangal SP; Antiulcer action of Leaves Extract of Murraya Koenigii In Experimentally induce Ulcer In Rats; Pharmacologyonline; 2011;2;818-824.
- 60) Thilahgavani N, Ramasamy P, Mohd Effendy A, Thirukanthan CS and Charles SV; Biological action of carbazole alkaloids and essential oil of Murraya koenigii touching antibiotic opposing microbes and cancer cell lines. Molecules; 2011;16;9651-9664.
- 61) Rani U, Verma N and Batra A; Total phenolic (flavonoid) contents and antioxidant capability of Murraya koenigii extracts International Journal of Medicinal Plant Research; 2012;1(7);124-128.
- 62) Bertha F, Lourdes P, Tricia P and Shruthi SD; Evaluation of Murraya extracts for calculating fungi development on zapota through inference studies; International Journal Of Pharmaceutical Sciences And Research; 2012;3(7);2196-2200.
- 63) Deepa I and Uma P; Radioprotective action of Murraya koenigii (L.) on cellular antioxidants in Swiss albino mice; Journal of Pharmacy Research; 2009;2(3);495- 501.
- 64) Biswa ND and Bishyajit KB; Analgesic action of leaf take out of Murraya koenigii;

International Journal of Comprehensive Pharmacy; 2012;4(2);1-3.

- 65) Vaibhav MD,Vijay RP and Amol BC; Antiinflammatory action of Murraya koenigii spreng on investigational animals; 2011;1(1);65-69.
- 66) Hassan Y, Qamar UA, Rahojoe IS, Seikh Farid UA and Norlewati AT; helpful cause of the trees of Murraya koenigii (Linn.) Spreng (Rutaceae) on diabetes- induced renal damage in vivo; Journal of Ethnopharmacology; 2011;135;88-94.
- 67) Kumar V, Angshu B, Sharma V, Suthar Sushil and Sunil T; injury Healing action of Murraya Koenigii in far above the ground Fat Diet And Streptozotocin Treated Type-2 Diabetic Rats; International Journal Of Research In Pharmacy And Science; 2011;1(2);128-140.
- 68) Debosree, Ghosh, Syed benazir F, Elina M, Monalisa D and Debasish B; protecting effect of aqueous leaf take out of Murraya koenigi against lead induce oxidative stress in rat liver, kindness and kidney: a dose response study Asian Journal of Pharmaceutical and Clinical Research; 2012;5(4);54-58.
- 69) Shamim Q, Mobeen S, Jitendra P, Kumar GS, Syed zia UH and Syed S; Comparitive Antihelmintic action of Aqueous and ethanolic take out of stem bark extract of Murraya koneigii; International Journal of Pharmaceutical Research And Development; 2011;3(5);96-101.
- 70) Dinesh KP, Narendra Y, Vinod N, Pradeep S and Ajay B; injury healing action of Murraya koenigii leaf extract; International Journal of Comprehensive Pharmacy; 2010;1(4);1-2.
- 71) Mohammed U and Barhate SD; Investigation and lessons of anti-inflammatory activity of Murraya koenigii spreng. Leaves;



International Journal of Pharmacognosy and Phytochemical Research; 2012;4(1);12-16.

72) Juhi M, Asia Y, Rattan deep S and Aradhana; phytochemical investigation and in- vitro antioxidant potential of leaves of Murraya koenigii; International journal of integrative biology; 2009;7(3);171-174. HOW TO CITE: Dr. Santhosh Suddagoni\*, Dr. kapil malviya, Dr. Santikari Sesha Phanindra, Dr. M. Venkata Ramana, Bodige Divya, Dr. Ganesh Akula, Nephroprotective and Anti-Inflammatory Activity of Combination of Punica Granatum (Fruit Peel) And Tectona Grandis (Bark), Int. J. in Pharm. Sci., 2023, Vol 1, Issue 6, 86-98. https://doi.org/10.5281/zenodo.8039301

