



**INTERNATIONAL JOURNAL OF
PHARMACEUTICAL SCIENCES**
[ISSN: 0975-4725; CODEN(USA): IJPS00]
Journal Homepage: <https://www.ijpsjournal.com>



Review Paper

Mass Spectrometry -A Powerful Tool for Modern Analysis

Ashwini Shinde*, Shruti Mali, Mrudula More

Eklavya College Of Pharmacy

ARTICLE INFO

Published: 14 Feb 2026

Keywords:

Mass, Charge, Velocity, Ion,
Fragment, Protein

DOI:

10.5281/zenodo.18640058

ABSTRACT

Mass spectrometry is a technique used to find and identify proteins and other molecules in biological samples. It can also help in studying how proteins interact with each other. In this method, a molecule is broken into charged pieces. These pieces (ions) are then moved, separated, and detected based on their mass-to-charge (m/z) ratio. The detector shows how many ions are present by measuring their signals. This chapter also explains the common problems that can occur while collecting raw data and how the data is processed to read the mass spectrum. First, peaks are selected from a survey scan. These peaks are then studied in MS/MS to find the mass of peptide fragments. The fragments are matched with a database to get a peptide score. Finally, proteins are identified from the correctly matched peptides

INTRODUCTION

Mass spectrometry (MS) is a powerful method used to study different chemicals in areas like the environment, medicine, pharmacy, forensics, and food testing(1). In this technique, the sample (solid or liquid) is changed into gas and then into charged particles called ions(2). These ions are separated based on how they move in electric and magnetic fields. The results appear as a mass spectrum, which is a graph showing different ions based on their mass(3). If a pure substance is present, the graph shows a strong peak with a high m/z value. When MS is connected with instruments like HPLC or GLC, it becomes more sensitive and can

detect even very small amounts of a substance(4). MS is also used to find the structure and molecular weight of organic and biological compounds. It is an important tool in proteomics, the study of protein structures (5).

PRINCIPLE-

In a mass spectrometer, the sample is first heated to create a high vapor pressure, which helps the molecules break into fragments and become ionized. These charged ions are then accelerated by applying voltage, making them move through the mass analyzer based on their mass. Ions that have the same charge travel with the same speed.

*Corresponding Author: Ashwini shinde

Address: Eklavya college of pharmacy ,tasgaon

Email ✉: ashwinishinde0627@gmail.com

Relevant conflicts of interest/financial disclosures: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.



Next, the ions enter a magnetic field in the detector. This magnetic force pushes ions with the same speed into a curved or circular path. Charged ions bend because the magnetic field acts like a centripetal force, pulling them toward the center. Only the ions that have the right combination of

electric and magnetic properties pass through without bending and reach the data system, where their signals are recorded. The mass spectrum created from this process is then used to study and identify substances in the sample(6)

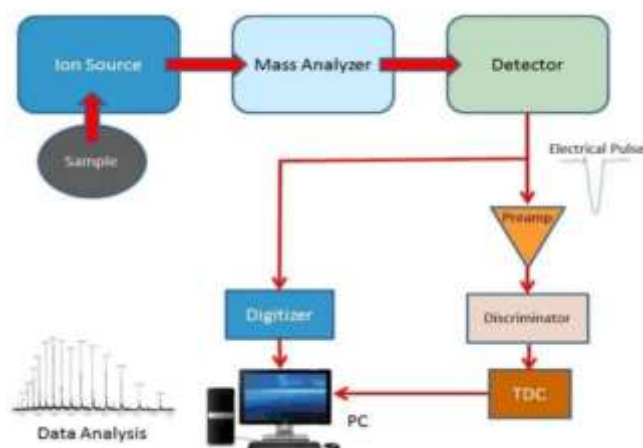


Fig no .1 working of mass spectroscopy

WORKING–

Before a sample can be analyzed in mass spectrometry, it must be properly prepared for ionization, which is an essential step in the working of the instrument. Samples are commonly converted into liquid or gaseous form using chromatography techniques such as gas chromatography and liquid chromatography(7). In gas chromatography, the sample is diluted, vaporized, and separated based on properties like molecular size, shape, boiling point, and weight, after which the volatile components are directed into the mass spectrometer for ionization and detection(8). Liquid chromatography separates components according to their interaction with the mobile and stationary phases, mainly influenced by polarity, allowing each compound to enter the mass spectrometer individually for further ionization, mass separation, and detection(9). Other ionization-based preparation methods, such as electrospray ionization and

fast atom bombardment, help convert samples into ions directly from liquid or solid phases(10). These steps enable the mass spectrometer to accurately analyze a wider range of biomolecules, including proteins, nucleic acids, lipids, and fatty acids, as part of its overall working mechanism(11,12).

RECENT ADVANCEMENT –

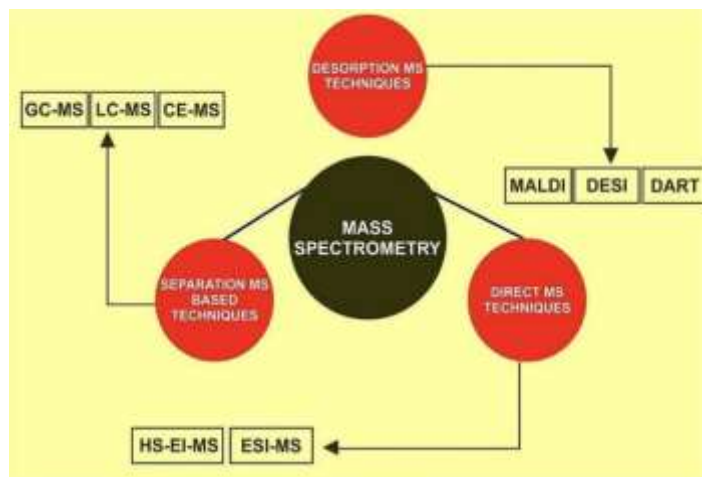


Fig no.2 Recent advancement of mass spectroscopy

APPLICATION-

[1] Analysis of Aerosol Particles-

- Helps monitor and control air pollution.
- Assists in predicting climate change effects.
- Evaluates health risks from airborne particles.
- Identifies sources of dust, smoke, and pollutants
- Supports environmental and atmospheric research (13,14,15).

[2] Emerging Medical and Biomedical -

- Detects age-related protein changes.
 - Identifies disease biomarkers (Alzheimer's, Parkinson's).
 - Detects antibiotic-resistant microbes.
- Used in toxicology and drug testing.
- Check herbal, nutrition, and supplement Composition
- (15,16,17,18,19,20,21,22,23,24,25,26,27)

[3] Identification of Environmental Pollutants

Mass spectrometry (MS) can be used to identify environmental pollutants such as nitrogen oxides, sulfur dioxide, plastics, lead, and particulate matter that are harmful to

both humans and animals. By combining the selectivity of liquid chromatography (LC) with the high sensitivity of MS, LC-MS has become a powerful tool for environmental analysis. It helps detect steroid estrogens and nitrosamines in wastewater, analyze polar, ionic, and heat-sensitive substances, identify antibiotics in pesticides, and detect perfluorinated organic (PFO) compounds found in cleaning products, textiles, and photographic materials. Thus, LC-MS plays an important role in assessing and controlling environmental pollution (28).

[4] Proteomics Mass spectrometry (MS) is widely used for protein analysis, helping in protein characterization and sequencing through soft ionization methods like electrospray and MALDI. It can identify proteins of different molecular weights, study changes in protein expression, detect post-translational modifications, and analyze important interactions such as protein–ligand, protein–protein, and protein–DNA interactions (29).

CONCLUSION

Advanced mass spectrometry techniques can now accurately detect and identify over 2,000 proteins within a single proteome. In complex mixtures, the major challenges are not sensitivity, but rather the wide dynamic range—where highly abundant peptides can overshadow those present in low amounts—and the limited sequencing speed of the instrument.

REFERENCES

1. H. H. Willard, L. L. Merritt, Jr., J. A. Dean, F. A. Settle, Jr. (1988) *Instrumental Methods of Analysis*, Wadsworth Publishing Co., Belmont, CA, pp. 465-507.
2. Sparkman, O. David (2000). *Mass spectrometry desk reference*. Pittsburgh: Global View Pub. ISBN 978-0-9660813-2-9.
3. J. Barker, *Mass Spectrometry, Analytical Chemistry by Open Learning*, Wiley, 2nd Edition (1999).
4. J. T. Watson, O. D. Sparkman, *Introduction to Mass Spectrometry—Instrumentation, Applications and Strategies for Data Interpretation*, 4th Ed., John Wiley and Sons, Chichester, 2007.
5. E. De Hoffmann, V. Stroobant, *Mass Spectrometry – Principles and Applications*, 3rd Ed., John Wiley and Sons, Chichester, 2007.
6. Strathmann FG, Hoofnagle AN. Current and future applications of mass spectrometry to the clinical laboratory. *Am J Clin Pathol*. 2011; 136: 609-616.
7. Bourgoigne E, Wagner M. [Sample preparation and bioanalysis in mass spectrometry]. *Ann Biol Clin (Paris)*. 2015 Jan-Feb;73(1):11-23. [PubMed]
8. Fothergill WT. Gas chromatography. Technique. *Proc R Soc Med*. 1968 May;61(5):525 -8. [PMC free article] [PubMed]
9. Rappold BA. Review of the Use of Liquid Chromatography-Tandem Mass Spectrometry in Clinical Laboratories: Part I-Development. *Ann Lab Med*. 2022 Mar 01;42(2):121-140. [PMC free article] [PubMed]
10. Meher AK, Chen YC. Electrospray Modifications for Advancing Mass Spectrometric Analysis. *Mass Spectrom (Tokyo)*. 2017;6(Spec Iss):S0057. [PMC free article] [PubMed]
11. Hemling ME. Fast atom bombardment mass spectrometry and its application to the analysis of some peptides and proteins. *Pharm Res*. 1987 Feb;4(1):5-15. [PubMed]
12. Züllig T, Köfeler HC. HIGH RESOLUTION MASS SPECTROMETRY IN LIPIDOMICS. *Mass Spectrom Rev*. 2021 May;40(3):162-176. [PMC free article] [PubMed]
13. Nash, David G.; Baer, Tomas; Johnston, Murray V. (2006). "Aerosol mass spectrometry: An introductory review". *International Journal of Mass Spectrometry*. 258 (1-3): 2-12. Bibcode:2006IJMSp.258....2N.
14. Pratt, Kerri A.; Prather, Kimberly A. (2012). "Mass spectrometry of atmospheric aerosols-Recent developments and applications. Part I: Off-line mass spectrometry techniques". *Mass Spectrometry Reviews*. 31 (1): 16.doi:10.1002/mas.20322
15. Pratt, Kerri A.; Prather, Kimberly A. (2012). "Mass spectrometry of atmospheric aerosols-Recent developments and applications. Part II: On-line mass spectrometry techniques". *Mass Spectrometry Reviews*. 31 (1): 17-48
16. Shuken, S.R., Rutledge, J., Iram, T. et al. Limited proteolysis-mass spectrometry reveals aging-associated changes in cerebrospinal fluid protein abundances structures. *Nat Aging* (2022).



17. Sebastián-Serrano Á, de Diego-García L, Díaz-Hernández M. The Neurotoxic Role of Extracellular Tau Protein. *Int J Mol Sci.* 2018;19(4): 998. Published 2018 Mar 27. Ozge Karayel, Sebastian Virreira Winter, Shalini Padmanabhan, Yuliya I. Kuras, Duc
18. Tung Vu, Idil Tuncali, Kalpana Merchant, Anne-Marie Wills, Clemens R. Scherzer, Matthias Mann. Proteome Profiling of Cerebrospinal Fluid Reveals Novel BiomarkerCandidates for Parkinson's Disease. *bioRxiv* 2021.07.22.453322.
19. Michno W, Blennow K, Zetterberg H, Brinkmalm G. Refining the amyloid β peptide molecular characterization in brain, cerebrospinal fluid, blood, and plasma. *J Neurochem.* 2021 Oct; 159(2): 234-257.
20. Hrabak J, Bitar I, Papagiannitsis CC. Combination of mass spectrometry and DNA sequencing for detection of antibiotic resistance in diagnostic laboratories. *Folia Microbiol (Praha).* 2020 Apr; 65(2): 233-243.
21. Charretier Y, Schrenzel J. Mass spectrometry methods for predicting antibiotic resistance. *Proteomics Clin Appl.* 2016 Oct;10(9-10): 964-981.
22. Liang T, Leung LM, Opene B, Fondrie WE, Lee YI, Chandler CE, Yoon SH, Doi Y, Ernst RK, Goodlett DR. Rapid Microbial Identification and Antibiotic Resistance Detection by Mass Spectrometric Analysis of Membrane Lipids. *Anal Chem.* 2019 Jan 15; 91(2): 1286-1294.
23. Abril AG, Carrera M, Böhme K, Barros-Velázquez J, Calo-Mata P, Sánchez-Pérez A, Villa TG. Proteomic Characterization of Antibiotic Resistance in *Listeria* and Production of Antimicrobial and Virulence Factors. *Int J Mol Sci.* 2021 Jul 29; 22(15): 8141.
24. Wu AHB. The Impact of Mass Spectrometry on Patients' Medical and Nonmedical Lives. *Lab Med.* 2021 Jul 1; 52(4): e58-e65.
25. Holzlechner M, Eugenin E, Prideaux B. Mass spectrometry imaging to detect lipid biomarkers and disease signature cancer. *Cancer Rep (Hoboken).* 2019 Dec; 2(6): e1229.
26. Haartmans MJJ, Emanuel KS, Tuijthof GJM, Heeren RMA, Emans PJ, Cillero-Pastor B. Mass Spectrometry-based Biomarkers for Knee Osteoarthritis: A Systematic Review. *Expert Rev Proteomics.* 2021 Aug; 18(8): 693-706.
27. Lădaru A, Bălănescu P, Stan M, Codreanu I, Anca IA. Candidate proteomic biomarkers for non-alcoholic fatty liver disease (steatosis and non-alcoholic steatohepatitis) discovered with mass-spectrometry: a systematic review. *Biomarkers.* 2016; 21(2): 102-14.
28. Bene J, Szabo A, Komlósi K, Melegh B. Mass Spectrometric Analysis of carnitine and its Esters: Potential Biomarkers of Disturbances in Carnitine Homeostasis. *Curr Mol Med.* 2020; 20(5): 336-354.
29. Chromatography and mass spectrometry for environmental analysis. <https://www.azom.com/article.aspx?ArticleID=12518>.
30. Dancik V. et al. De novo peptide sequencing via tandem mass spectrometry. *J Comput Biol.* 1999; 6: 327-342.
31. M. Karas and F. Hillenkamp, Laser desorption ionization of proteins with molecular weight exceeding 10,000 daltons, *Analyt. Chem.* 60 (1988), 2299-3201.

HOW TO CITE: Ashwini Shinde*, Shruti Mali, Mrudula More, Mass Spectrometry -A Powerful Tool for Modern Analysis, *Int. J. of Pharm. Sci.*, 2026, Vol 4, Issue 2, 2299-2303. <https://doi.org/10.5281/zenodo.18640058>

