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Research Paper

Formulation & Evaluation of Herbal Tablet on Anti-Cancer

D. N. Shinde*, V. J. Zanwar, A. S. Kale

Rajashree Shahu College of Pharmacy, Markhel, Maharashtra, India

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ABSTRACT

The present study was designed to develop and evaluate herbal anti-cancer tablets through the selected bioactive compounds from plant origin. For the preparation of this tablet, herbal extracts including Lantana camara along with pharmaceutical excipients such as lactose, starch, talc magnesium stearate, acacia vanilla and methyl paraben were used. Direct compression method was adopted for preparation of the formulation it is generally used method for the production of solid dosage forms, as compared with other methods is easier to prepare and also cheaper. Some of the well-known medicinal plants were elected for this study on the basis of their anticancer potential like Curcuma longa, Withania somnifera and Camellia sinensis. The herbal extracts were prepared by appropriate method of extraction and were formulated into tablets along with other suitable excipients to ensure stability its compressibility and satisfactory pharmaceutical properties. By assessing pre-compression characteristics such angle of repose, bulk density, tapping density, Carr's index, & Hausner ratio for the manufactured granules, the powder mixes flowability and compressibility were evaluated. Following tablet preparation, thickness, hardness, friability, weight variation, and disintegration time were assessed. Alkaloids, flavonoids, tannins, phenolic compounds, saponins, and other biologically active chemicals with anticancer action were found in the extract after initial phytochemical testing was carried out as previously mentioned. Additionally, cancer cell lines like HeLa and MCF-7 can be used to evaluate the anticancer activity of the prepared herbal remedy. All the evaluation parameters was at a range that were acceptable based on pharmaceutical limitations indicated which implies the preparation herbal anti-cancer tablets republished in range. The formulation exhibited good physical properties and sufficient mechanical strength, which may justify its potential as a herbal anticancer dosage form.

INTRODUCTION

IntroductionMedicinal plants have traditionally been a relevant source of pharmacologically active agents for the prevention and treatment of

***Corresponding Author:** D. N. Shinde
Address: *Rajashree Shahu College of Pharmacy, Markhel, Maharashtra, India*
Email ✉: dnyaneshwarshinde1032@gmail.com

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diseases2. Plant-origin medicines were used for the management of several health ailments in traditional medicine systems since ancient times. This systematic and scientific approach of understanding about medicinal plants offers an enormous repertoire of bioactive phytoconstituents which may be used as lead molecules for the discovery of modern pharmaceuticals. Recent advances in a tool called the scientist's pipeline have enabled researchers to find and classify large volumes of biologically active compounds from plants. Many plants were reported to have anticancer, antiinflammatory, antidiabetic, antimicrobial, antifungal, hepatoprotective and antioxidant as well as anthelmintic activities. Lantana camara Linn., a blooming ornamental shrub related to the Verbenaceae family, is an example of a medicinal plant. The plant is known by several names, includes West Indian panama, Spanish flag, wild sage, and Surinam tea plant. Lantana camara is frequently employed in herbal therapy to treat a variety of illnesses. Among the traditionally used medicinal plants, turmeric is one of the most making it significant in modern pharmacotherapy due to its diversity of biologically active phytochemicals. Cancer is a serious problem of global health and at present become 2ndleading cause of death across the globe. The incidence rate of cancer in general also continues to increase and is expected to especially explode in the coming decades. Cervical cancer based on the various types of malignancies is one of the most common cancers affecting women, and prostate cancer is considered one of the most frequent solid tumors in men. Conventional cancer therapies such as radiation therapy and chemotherapy are expensive and cause unwanted side effects. It is these shortcomings that have led to a significant interest in exploring naturally sourced phytocompounds as viable therapeutic agents. The anticancer activities of plant extracts and essential oils compounds ascribed to their bioactive content, capable of tumor growth inhibition or cancer cells apoptosis were illustrated in many studies. Numerous cancer cell lines are sensitive to the diverse cytotoxic activity of essential oils and plant aromatic volatile compounds. The activity of Lantana camara in particular, antimicrobial, antioxidant and cytotoxic activity has been extensively investigated. The predominant pharmacological properties imply that plant may hold extensive therapeutic values in designing new herbal anticancer formulations.[1,2]

PLANT DESCRIPTION:

Lantana camara is a low-growing, upright or semi-scandent shrub with a tetragonal stem, strong, recurved prickles, and a black current-like scent. Plant Size: 2.5 m in spread, 1-3 m in height. Ovate to ovate-oblong leaves with a serrated tip might be acute or subacute. Both the two sides of leaves are rugose, with tiny hairs covering both sides. The simple, opposite, green leaves are 3–8 cm lengthy and 3–6 cm broad.

The plant features tightly clustered small blooms known as umbels. They are available in a range of colors, including orange, red, yellow, or white. Significantly, these flowers develop color as they mature. The yellow-throated flowers, produced in axillary heads most of the year.

Calyx small; corolla tube slender with lobes spreading c. 6–7 mm in diameter, unequally divided. Further leaves arise from a whorl of four stamens in two pairs with bicarpellary ovary and two ovules.

Inflorescences are globose to ovoid, 2–3 cm in diameter and contain 20–40 sessile flowers.

An extensive root system Enables Lantana camara to emerge new growth after repeated cutting and therefore some vigorous regrowth and a pattern of widespread localization.[3]

GEOGRAPHICAL DISTRIBUTION:

The tropical plant species Lantana camara, sometimes known as widespread lantana, is native to the Caribbean and Central and Northern South America. Due to its rapid expansion, the plant now exists in almost sixty nations, including Brazil, Mexico, Florida, New Zealand, Trinidad, and Jamaica. The plant is indigenous to numerous African nations, including Uganda, Kenya, Tanzania, and South Africa, in addition to Madagascar. Lantana camara is reported to have been introduced in India prior to the 19th century and widely disseminated across regions of the country.

In Indian languages, the plant has different names such as Raimuniya (Hindi), Chaturangi or Arippu (Sanskrit)), Konginipoo (Malayalam), Tantani and Ghaneri (Marathi), Pulikampa (Telugu) & Natahu (Kannada).[4,5]

TAXONOMY:

Kingdom: Plantae
Division: Magnoliophyta
Class: Magnoliopsida
Order: Lamiales
Family: Verbenaceae
Genus: Lantana
Species: Lantana camara[1,4]

Fig. 1: Morphological features of Lantana camara depicting flowers and fruits

AIM:

Preparation and evaluation of herbal anticancer tablet from Lantana camara leaf extract

OBJECTIVES:

1. To obtain Lantana camara extract to make an herbal tablet.
2. To see if the formulation itself can halt or slow the growth of cancer cells.
3. In an effort to see if they could make a brew that would reduce cancer progression risk.
4. To develop a cost-effective herbal tablet.

LITERATURE REVIEW:

Lantana camara has been documented to have pharmacological activities and therapeutic potentials Plant has many biologically active constituents which found responsible for various types of medicinal activities as results various phytochemical and pharmacological studies

- Poonagithai A. (2013) studied the presence of important phyto-constituents viz: alkaloids, saponins, tannins, carbohydrates and proteins through qualitative and quantitative phytochemical analysis on Lantana camara leaves using various target compounds. The constituents of these compounds are suspected to play a role in the biological activity variations of the plant.
- Lantana camara Extracts' In-Vitro Anticancer Activity and Initial Phytochemical Analysis According to study done by Babar V. B., Khapale P. R., and Nagarale S. N., flavonoids and other bioactive compounds may be the

cause of the plant extract's cytotoxic action on lines of cancer cells.

- A thorough analysis of Lantana camara's beneficial properties was published by Kalita S., Kumar G., Karthik L., and Rao K.V. In to their research, the plant possesses several health benefits, including antibacterial, anti-inflammatory, antioxidant, and anticancer properties.
- 11 Herbal anticancer tablet formulation and evaluation of its by Madhekar S.M., Gandhari A.R., Tidke V.N., Pawar R.G. and Lanjewar S.H. Preformulation studies were carried out in their investigation along with evaluation parameters such as angle of repose, hardness and friability, fir tablets; quality control tests
- Mishra A. and Mishra S.: Development and Evaluation of Antifungal Polyherbal Soap Containing Lantana camara Extract. The study was important in producing meaningful information on extraction method and utilization of plant extracts for pharmaceutical formulation.

Lantana camara has been recognized as a significant source of all various types of medicinal usages which can be studied for their pharmacological potential.[6-11]

MATERIALS AND METHODS:

a. Plant Material

Fresh Lantana camara leaves were gathered from the neighborhood. Nanded Markhel (Maharashtra, India). Plant Material Isolation, Verification, and Application in Additional Research Procedures

b. Preparation of Plant Extract

Before being rinsed with distilled water, leaves were first cleaned with tap water to get rid of dust and other impurities. Finally, the (Inf) particle plant material was shade-dried for two days at 30 °C to completely remove any remaining moisture. After being completely dehydrated, they were ground into a powder with a mortar and pestle. Kazemi et al. 201231 About 20 g the dried powdered plant material was immersed in an appropriate volume of methanol solvent in a vacant container as left for three to four days to allow for the palatable recovery of plant-based substances.

The extract was decanted and filtered with adequate means of filtration. This extract was filtered out, placed in petri dish and dried at room temperature for 2 days to obtain the plant extract.[12,13]

FORMULATION OF HERBAL TABLET:

Table 1: Composition of Lantana camara Extract Tablet Formulation

Sr. No.	Ingredient	Quantity	Role
1	Lantana camara extract	2.5 g	Active ingredient
2	Lactose	2.2 g	Diluent
3	Gum Acacia	0.075 g	Stabilizer / Binder
4	Starch	0.125 g	Binding agent
5	Magnesium stearate	0.05 g	Lubricant

Tablet Preparation Method

The Direct Compression Method

The technique most commonly employed to create solid dosage forms in the pharmaceutical sector, direct compression, has been employed to make the tablets.

Here is the process:

- Weighing – The amount of active ingredient as well as excipients were accurately weighed in using a balanced scale.
- Sieving – All the Ingredients were sieved through sieve No. 16 to pass bulk homogenouse particle size.

- It prepares/mixes the elements well till a unique and concentrated material or subbiocomposite is obtained, which is executed with mortar and pestle to allow for feature systematic distribution.
- Compression – Tablets were prepared using a tableting punching machine, the homogeneous powder mixture was compressed.[14]

Figure 2: Final Tablet Formulation of Lantana camara Extract

PREFORMULATION STUDIES:

a. Organoleptic Properties:

Visual evaluation of organoleptic characters of Lantana camara powder

Table 2: Organoleptic Properties of Lantana camara Extract

Sr. No.	Parameter	Observation
1	Colour	Greenish
2	Odour	Mucilaginous
3	Taste	Bitter

Figure 3: dissolving the plant extract

b. Solubility:

The plant extract was dissolved in different solvents (water, methanol and ether) to determine its solubility.

Table 3: Lantana camara Extract Solubility Profile

Solvent	Solubility
Water	Completely soluble
Methanol	Slightly soluble
Ether	Very poorly soluble

c. pH Determination:

The extract of plant was measured with the digital pH meter. Instrument was calibrated at acetate buffer solution before measurement. Its pH was 5.8.[10]

d. Angle of Repose:

The properties of the powder blend are examined using the degree of repose. A conical pile of powder is formed by dropping the powder via a funnel onto a level surface. The formula as follows was used to get the angle of repose:

$$\theta = \tan^{-1}\left(\frac{2h}{d}\right)$$

Where

h = height of powder pile

d = diameter of the base

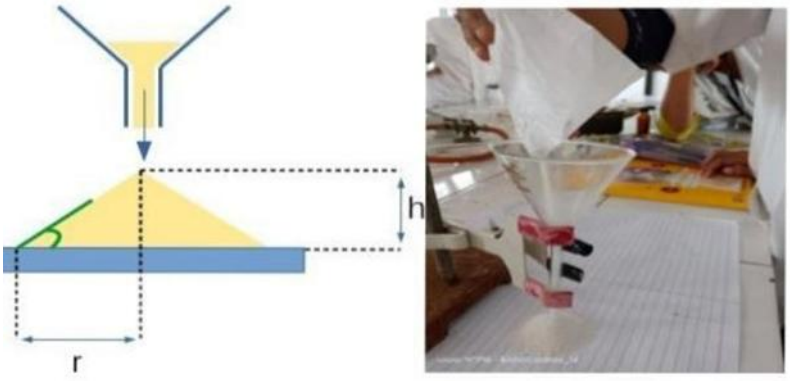

Figure 4: Determination of Angle of Repose of Powder

Table 4: Classification of Powder Flow Properties by Angle of Repose [15]

Flow Property	Angle of Repose
Excellent	25–30°
Good	31–35°
Fair	36–40°
Passable	41–45°
Poor	46–55°

EVALUATION OF TABLETS:

a. Size and Shape:

It is dimensional, measurable and controllable. An only variable that has measured in micrometer or another device, tablet thickness. For tablet thickness control, it should be +/−5% variation of standard value. [16]

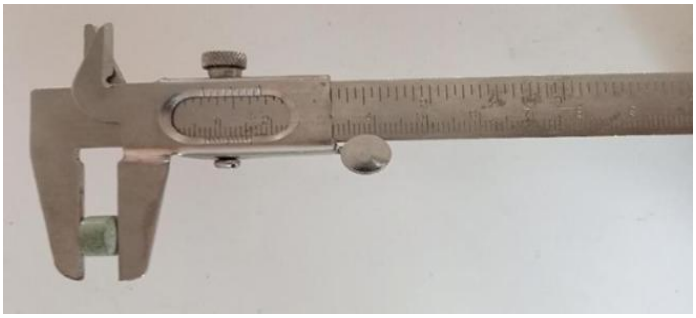

Fig 5: Measuring Thickness of Tablet with Vernier Caliper

b. Hardness Test:

In terms of tablet hardness, this is the diameter compression force which compensates for breaking a tablet. To withstand the mechanical shock of handling during creation, packing, and shipping, tablets have to have adequate hardness or strength and resistance to friability. Hardness is often used to gauge a tablet's crushing power. How

may the power of the pill be obtained? Use the thumb as a base to smash the tablet between the your two remaining fingers. A rapid breakage is a sign that the tablet is strong enough. A tablet is placed between two anvils in the strength tester, and force is applied to these two anvils to determine the crushing strength, which can break or destroy tablets. They are used as hardness testers. 1. The Monsanto The tester 2. The Strong-Cobb Tester 3. The Pfizer Tester 4. Tester Erwika Fifth. Schleuniger Tester Hardness compressed tablet: 5 — 8 kg. [17]

Figure 6: Determination of Tablet Hardness Using Monsanto Hardness Tester

c. Friability Test

Outcomes Tablets that powder and crumbles upon handling. Non-elegant and consumer acceptance Put really dirty processes into production components Affect on weight variation of tablet or content uniformity as well. Operating concept for the laboratory friability test: The Roche friabilator is used to conduct the malleability test. They are physically dropping a foot through the friabilator which runs for 100 revolutions, because they are made of a plastic chamber that the revolves at 25 rpm. We reassess the medicine. A drop of <0.5 to 1.0% tablet is in limits on average up to October 2023.[18]

$$\%Friability = \frac{W_0 - W_f}{W_0} \times 100$$

Where,

W_0 = Initial weight of tablets

Wf = Final weight of tablets

Acceptable tablets recorded less than 1% weight loss.

Figure 7: Determination of tablet friability by using friability tester

Table 5: Friability Test Result of Lantana camara Tablets

Tablet name	Initial weight	Final weight	Weight variation (%)
Lantana camara tablet	5.50	5.00	0.50

d. Weight Variation Test (USP):

The United States Pharmacopeia (USP) standards for tablet capacity homogeneity were followed when conducting the mass variation test. An analytical balance (Shimadzu AY220, Kyoto, Japan) was utilized to weigh each of the twenty randomly chosen pills. After calculating the average weight of ten tablets, each tablet's weight was contrasted to the standard.

USP: The percentage within limits (not greater than two units exceed the prescribed value, and no unit depart from more than double numbers). Permitted weight variations are based on the average weight of the tablet.[19]

Weight Variation Limits:

Table 6: Weighing Variation Limits for Tablets according to Pharmacopoeia

Sr. No.	Average Weight of Tablet (mg)	Maximum % Difference Allowed
1	130 mg or less	10%
2	130–324 mg	7.5%
3	More than 324 mg	7%

Calculation of Weight Variation:

The percentage change in weight was given by the following equation:

$$\text{Weight Variation (\%)} = \frac{(IW - AW)}{AW} \times 100$$

Where:

IW = Weight of single tablet

weight_avg = Average weight of tablets

Observed Weight Variation Data:

Table 7: Change in weight of Lantana camara tablets

Individual Weight of Tablet (g)	Difference from Mean	(%) RSD
0.57	0.02	3.6
0.54	-0.01	-1.81
0.61	0.06	10.9
0.56	0.01	3.7
0.57	0.02	3.6
0.55	0	0
0.50	-0.05	-0.09
0.48	-0.07	-12.7
0.57	0.02	3.63
0.54	-0.01	-1.8

Total % RSD = 4.3

e. UV–Visible Spectroscopy:

An analytical technique called UV-visible spectroscopy may be used to identify chemical species based on how they absorb visible or ultraviolet light, which results in wavelengths that form different spectra. This is done based on the interactions of electromagnetic radiation with matter. In this approach, the compounds absorb radiation in ultraviolet and visible electromagnetic spectrum. The range of 200–400 nm is called as ultraviolet region and that from 800-400 nm belongs to the visible region. The simple inhibition form is valuable in chemical qualitative and quantitative structure analysis.[20]

Figure 8: UV–Visible spectrophotometer

To prepare the 0.1 M sodium hydroxide solution:

a clean, dry 1000 ml volumetric flask was filled wit around 100 ml of water that had been distilled. Stirring until everything was dissolved, 4 g of potassium hydroxide was added evenly. A a solution of 0.1 M sodium hydroxide, was prepared by adding pure water to the final volume of 1000 ml after the solute had completely dissolved.

To make a 0.1 M sodium hydroxide solution:

fill a clean, dry 1000 ml flask with with 100 ml of distilled water. The increment addition of ~ 4 g of sodium hydroxide was added with vigorous stirring until it completely dissolved. A final 0.1 M sodium hydroxide solution was prepared by filling up to the desired volume with distilled water, and this was repeated three times (dilution step) after the solute was dissolved in distilled water.[21]

Procedure:

1. A mortar and pestle were used to weigh and break the pills into a finely ground powder. For analysis, a weighed quantity of powdered specimen—equivalent to 0.15 g of anti-cancer tablets—was determined.
2. A dry 200 ml flask was taken and funneled.
3. The weighed amount of powder was then transferred to the funnel and subsequently into a volumetric flask.
4. After that, 50 ml of the 0.1 M sodium hydroxide solution was add through funnel to wash down any sample remaining at the inside walls of flask into flask.
5. To guarantee dissolution, the mixture was agitated for 15 minutes until being brought to a final volume of 100 milliliters with distilled water.
6. Next, the distilled water was added to the mix until it reached a volume of 200 ml.
7. Whatman filter paper was used to sieve this prepared solvent to get rid of any elements that weren't dissolved.
8. ten milligram of the filtrate were mixed with one hundred milliliters of distilled water.
9. A fresh 100 ml volumetric flask was filled with an additional 10 ml of this solution, that was diluted by adding a 0.1 M sodium hydroxide in order solution.
10. To achieve a final volume, the solution was diluted with distilled water and well stirred.
11. The produced solution was scanned at 257 nm using a UV-visibleA (1%, 1 cm) =715 at257 Drug content was calculated.[12]

f. Disintegration Test (USP):

In this study, the disintegration test was performed as per United States Pharmacopeia (USP) guidelines to determine the time taken for tablets to break down into clumps of smaller particles given under certain conditions. The USP breakdown equipment, for for example, consists of six clear tubes that are about three inches long, open at the highest point, and screened by a 10-mesh screen.

For this test, one tablet was placed in each of two tubes that were then positioned bottom up using a basket-type rack assembly and immersed in either 1 liter of disintegration medium (distilled water, simulated gastric fluid or simulated intestine depending on experimental setup) which through out the experiment was maintained at 37 ± 2 °C simulating physiologic conditions.

During the test, the basket rack assembly moved vertically within a range of 5 to 6 cm at an orbital speed from 28 to 32 times per minute. They measured how long it took for the tablets to completely break down into particles small enough to fit through a mesh screen. Disintegration times acceptable for uncoated tablets generally fall within the range of 5 to 30 min.[18,19]

Figure 9: Disintegration test apparatus

g. Dissolution Test:

The process by which a solid drug material dissolves into a solvent and forms a solution is known as the process of dissolution. The technique of identifying the rate and degree of drug release from a dosage form under circumstances favourable to maximal controlling is known as dissolution testing, and it is a crucial metric for assessing bioavailability (Stier & Chernick, 2016). Standard USP equipment, often known as type I (basket type) and type II (paddle type), was used to conduct the United States Pharmacopeia (USP) dissolution test. This method involved placing a tablet in a web of wires basket that was fastened to the bottom of a revolving shaft driven by an engine with an adjustable speed.

A phosphate buffer served as the dissolution medium into which a tablet-containing basket was dipped inside a dissolution vessel. In a thermostatically controlled water bath, temperature of the medium was kept constant on 37 ± 5°C to better mimic physiological conditions. This process was carried out at a fixed rotating rate of the shaft to allow contact between the dissolution medium and the tablet's surface. At fixed time intervals, a sample was collected to measure the amount of drug released into the dissolution medium as function of time.[16,17]

Figure 10: Dissolution test apparatus

RESULTS:

Table 8: Preformulation Studies of Lantana camara Powder

Sr. No.	Parameter	Observation
1	Colour	Greenish
2	Odour	Mucilage
3	Taste	Bitter
4	Solubility	Completely soluble in water; slightly soluble in methanol
5	pH	5.8
6	Angle of Repose	28°

Table 9: Evaluation of Herbal Anticancer Tablet

Sr. No.	Parameter	Observation
1	Hardness	5 kg/cm²
2	Size and Shape	10 mm, spherical
3	Friability	0.5%
4	Weight Variation	3.5%
5	Dissolution Time	20 minutes
6	Disintegration Time	10 min 10 sec

CONCLUSION

Tablets were produced for the first time on the basis of Lantana camara leaf extract which was ascertained using direct compression method in this study as herbal anticancer tablets. The physicochemical and pharmaceutical attributes of the produced tablets were acceptable. Appropriate powder properties (a good angle of repose, for example) are indicated in preformulation studies (good flow properties during tablet manufacture). Following the compression, evaluation tests were conducted, and the outcomes, including hardness, friability, density variance, disintegration period, and dissolution time, were determined to be within the proper limits.The results suggest that prepared formulation of herbal tablet had appropriate mechanical strength, stability and pharmaceutical quality. Therefore, Lantana camara may serve as a potential natural source for preparing economical/ herbal anticancer formulations.

REFERENCES

1. Tabassum N and Vidyasagar GM: Antifungal investigations on plant essential oils: A review. International Journal of Pharmacy and Pharmaceutical Sciences 2013; 5(2): 19-28.
2. Bhuvaneswari E and Giri RS: Physicochemical and phytochemical screening of *Lantana camara* leaves. Journal of Pharmacognosy and Phytochemistry 2018; 7(6): 1962-1966.
3. Babar VB, Khapale PR and Nagarale SN: Preliminary investigation and in-vitro anticancer activity of *Lantana camara* L. (Verbenaceae). Journal of Pharmacognosy and Phytochemistry 2019; 8(5): 2524-2527.
4. Kalita S, Kumar G, Karthik L and Rao KV: A review on medicinal properties of *Lantana camara* Linn. Research Journal of Pharmacy and Technology 2012; 5(6): 711-715.
5. Barik SS, Sahoo RP and Barik SS: *Lantana camara* L.: An emerging threat to native flora and livestock. Journal of Pharmacognosy and Phytochemistry 2020; 9(5): 2363-2366.
6. Samarakoon SR, Ediriweera MK and Wijayabandara MD: Evaluation of anticancer activity of medicinal plants used in traditional medicine. Journal of Ethnopharmacology 2014; 155: 1331-1340.
7. Sriwiriyajan S, Tedasen A and Lailerd N: Anticancer activity of plant extracts against human cancer cell lines. BMC Complementary and Alternative Medicine 2014; 14: 432.
8. Nasr FA, Noman OM and Alqahtani AS: Cytotoxic activity of medicinal plant extracts against cancer cell lines. Saudi Pharmaceutical Journal 2018; 26: 815-822.
9. Omara EA, Nada SA and Farrag AR: Anticancer activity of plant-derived

compounds and extracts. Journal of Applied Pharmaceutical Science 2020; 10(6): 140-147.
10. Buranrat B, Suwannalert P and Srisawat T: Cytotoxic and apoptosis-inducing activity of plant extracts in cancer cells. Asian Pacific Journal of Cancer Prevention 2020; 21(6): 1675-1682.
11. Bayala B, Bassole IH and Gnoula C: Chemical composition and anticancer activities of essential oils from medicinal plants. Molecules 2020; 25: 1-16.
12. Diaz MM, Vivas JG and Hernandez F: Evaluation of medicinal plants with anticancer potential. Pharmacognosy Reviews 2022; 16(31): 1-10.
13. Niksic H, Kovac-Besovic E and Makarevic E: Chemical composition and biological activity of plant essential oils. Journal of Essential Oil Research 2021; 33(3): 197-205.
14. Ruttanapattanakul J, Wikan N and Smith DR: Biological activities of essential oils from medicinal plants. Molecules 2021; 26: 1-18.
15. Yeh HY and Lin YL: Bioactive compounds from medicinal plants and their pharmacological activities. Evidence-Based Complementary and Alternative Medicine 2021; Article ID 5571284.
16. Khalil MI, Ahmmed I and Ahmed R: Anticancer properties of plant-derived compounds: A review. Pharmacognosy Research 2021; 13(1): 1-8.
17. Udappusamy S, Sundararaj S and Chandrasekaran M: Antioxidant activity of *Lantana camara* leaf extracts. International Journal of Pharmaceutical Sciences and Research 2022; 13(2): 567-574.
18. Barros LM, Duarte AE and Morais-Braga MF: Chemical composition and cytotoxic activity of *Lantana camara* essential oil. Industrial Crops and Products 2016; 85: 150-158.
19. Bray F, Ferlay J and Soerjomataram I: Global cancer statistics. CA: A Cancer Journal for Clinicians 2018; 68(6): 394-424.
20. Gandaglia G, Leni R and Bray F: Epidemiology and prevention of prostate cancer. European Urology Oncology 2021; 4(6): 877-892.
21. Puyol M, Pujol JL and Gervais R: Advances in cancer therapy and global cancer burden. Cancer Treatment Reviews 2021; 95: 102179.

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